

## Using dissolved oxygen, pH, EC probes (for water quality monitoring)

There are different media which can be used for aquatic monitoring. The quality of water and particulate matter is determined by physical and chemical analysis, whereas living organisms can be used for biological water quality monitoring. The choice of the water quality variables will depend on the objectives of the programme, the occurrence of the variables (it has no use to produce, year-after-year, lists with “non- detectable”), the potential impacts (toxicity) and financial resources. Water used for irrigation will require quite different monitoring variables than water used for recreational purposes or drinking water extraction. A short overview will be presented here of the different water quality variables and they will be discussed further during the lecture:

- **Temperature:** a basic parameter, important for all chemical and biological processes. Large fluctuations are to be expected in deep lakes and reservoirs.
- **Colour:** is influenced by natural (e.g., humic acids) and anthropogenic sources; it may be important in view of the aesthetic quality of the water.
- **Odour:** is often caused by decomposition of organic compounds yielding organic acids, sulphides, etc. Odour is often an indication for bad water quality conditions.
- **Total suspended solids (TSS):** may especially be increased in rivers ("turbidity") during storm floods; TSS may carry the large bulk of micropollutants.
- **Electrical Conductivity (EC):** this easily determinable parameter presents a good measure for the total ions present in the water; in many cases there is a good correlation with the NaCl concentration.
- **pH:** just as temperature a basic water quality parameter, of importance to virtually all biological and chemical processes is pH. In most natural waters, pH values will be between 6 and 8.
- **Dissolved oxygen (DO):** is an essential component for all aquatic life cycles; concentrations < 2 mg O<sub>2</sub>/L will lead to deaths of most fish species. The theoretical maximum levels, i.e., solubility (ca. 10 mg O<sub>2</sub>/L) will largely be exceeded during daytime in eutrophic lakes and reservoirs. At higher temperatures the O<sub>2</sub> solubility will show a clear decrease. DO is mostly measured with a DO probe, but there is also a laboratory procedure ("Winkler titration").
- **Degradable organic matter:** high concentrations are due to wastewater discharges. As mentioned before, the term can be expressed as COD or BOD: Chemical and Biochemical Oxygen Demand, and is thus related to the dissolved oxygen levels.
- **Other inorganic variables** such as sulphides, silica, fluoride and boron should be monitored in case of serious problems. The same holds for arsenic, a major problem in many groundwaters.
- **Heavy metals (Pb, Cd, Zn, Hg, Cu, etc.):** usually these components are not part of basic monitoring programmes, because of the complexity of analysis in the often low concentration ranges encountered in practice. The dominant fraction is found in the particulate phase; therefore, determination in lake sediments often yields interesting results.
- **Organic micropollutants** such as PCBs (poly-chloro-biphenyls), PAHs (polycyclic aromatic hydrocarbons), and pesticides (DDT, aldrin, dieldrin, atrazine, etc.) can only reliably be measured in advanced monitoring programmes. These

micropollutants, together with the heavy metals, have a strong tendency for bioaccumulation in the food chain.

What is water quality?

Water quality is a measure of water's suitability to be used for a specific purpose, such as swimming, farming, or power generation. Water that is considered unsuitable for one application may be perfectly acceptable for another purpose. Quality is a statement of the physical, biological, and chemical characteristics of water based on key conditions. These conditions can vary by location, such as at different points in a river or by time depending on the climate. Surface water and ground water can also affect the quality of each other, since these two are connected at the water table. It is important to recognize that water quality can be adversely impacted by both natural and man-made factors. Regularly monitoring water sources can help identify potential issues before they cause serious harm.

Calibrating in the Field

Calibration is an essential step to getting accurate and repeatable results. Ideally, you should calibrate your testing equipment prior to use on the day of sampling. A post calibration check at the end of the day can be performed to determine if the instrument drifted out of calibration. Depending on the nature of your project, you may need to perform more frequent calibration checks throughout the day in addition to the check at the end of the day. Remember to only use fresh standards and clean probes as debris could adversely affect your results.

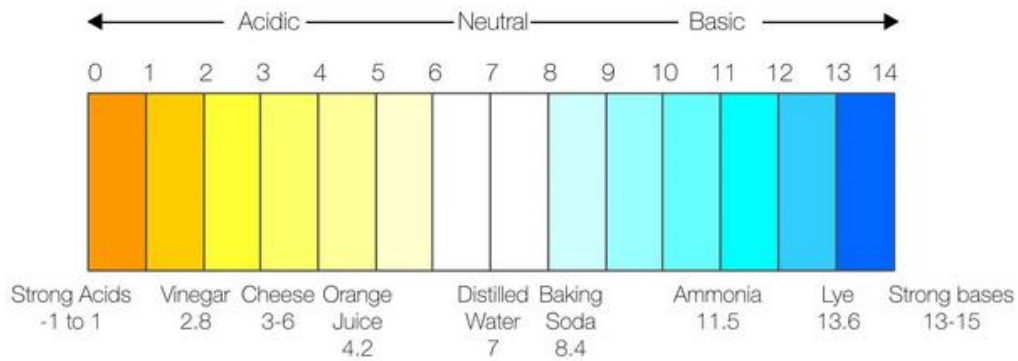
What are key water quality testing parameters?

There are a number of parameters that can be measured to indicate water quality. These parameters can be a measure of physical characteristics such as **pH**, conductivity, or temperature; a statement of the levels of various nutrients in water, such as nitrates and phosphates; or an indication of key elements and compounds in water, such as **dissolved oxygen**. Each parameter has some general standards and guidelines for determining if a tested sample should be considered acceptable or hazardous. The results of these tests are not necessarily absolute, since they must be compared in relation to what is considered normal levels for a body of water.

pH

### **What is pH?**

pH is a measurement of the relative concentration of hydrogen ions and hydroxide ions in water. The scale ranges from 0 to 14, with 0 being a strong acidic solution and 14 being strongly basic.



## Why is measuring pH important?

pH is a way to evaluate the suitability of water for living plant and animal organisms. If water has become too acidic or basic due to natural or man-made pollutants, there can be a profound negative impact on aquatic life. pH is considered normal in a body of water if it has a value of 5.0 to 9.0, but ideally it would fall in the range of 6.0 to 8.0.

## How do we measure pH?

Common pH tests, like chemical test kits and pH strips, are simple and inexpensive. However, they come with some issues that could lead to inaccurate results. Both of these methods of testing pH give you results based on a chemical reaction that results in a color change. When your paper or liquid sample changes color, you match it to the color guide provided and get your pH reading.

A more accurate means of testing pH is using a pH meter. When choosing a pH tester or meter, there are a number of considerations related to both the electrode as well as the device. Be sure to find a pH meter and electrode that is best suited for field work.

## pH Calibration in the Field

The first thing is to choose the buffer solutions that will bracket your expected value. What is bracketing? Also known as a two-point or multi-point calibration, bracketing consists of calibrating to two pH points – one above and one below your desired pH range. If the pH of your water sample is unknown, then a third calibration point will ensure the best accuracy.

### Calibration Procedure

1. Fill a beaker with enough pH calibration buffer to cover the electrode junction (about 75 mL in a 100 mL beaker).
2. Place the electrode in the beaker containing pH calibration buffer and gently stir.
3. Confirm the calibration point when the reading is stable, or when the digits do not change for at least 5 seconds.
4. Repeat for additional calibration points. Be sure to rinse with pure water between calibration points. At least two calibration points are recommended.

5. The calibration is complete. Rinse the probe with deionized water and store the probe according to manufacturer's instructions.

Temperature

## **What is temperature?**

Temperature is one of the most common measurements in our everyday life. In the context of water quality, temperature can provide an indication of the living conditions for aquatic plants and animals. Warm temperatures are generally considered beneficial for growth of aquatic populations. However, after a certain point temperature can have the opposite effect, contributing to a decline in the biological diversity in a water body.

## **Why is measuring temperature important?**

Aquatic organisms such as fish and plankton are cold-blooded, so the temperature of water has a direct impact on their body temperature. These organisms have ranges of temperature in which they can survive, or thrive. As the temperature reaches the high limit of its range for an organism, biological activity will be at a peak. This activity will decrease at the bottom of the range. If the temperature exceeds the acceptable range for an organism, the available supply of oxygen may be too low to sustain life. This is because warm water has an oxygen saturation point much lower than cold water. If temperature is below the acceptable range, not enough activity takes place to grow the species. High temperatures also contribute to the growth of algal blooms. Oxygen is consumed as these blooms are decomposed by bacteria, thus reducing the available dissolved oxygen supply.

Temperature in a water body varies based on the time of day and the amount of sunlight heating the surface of the water. Acceptable temperatures also vary depending on the type of river or stream being monitored. This depends on the watershed feeding the stream. If the stream is fed by a mountain spring, for example, the natural temperature of the stream may be quite cool (less than 68 degrees F). A stream that is considered warm water will have an average temperature greater than 68 degrees F but less than 89 degrees F. Temperature can also be influenced by the flow rate of a body of water. If the flow of water is increased, perhaps as a result of a heavy rainfall, the temperature can be expected to decrease. The increased current has a cooling effect on the temperature of the water.

Temperature pollution, also called thermal pollution, can be caused by runoff of water heated on asphalt or concrete. It can also be from industrial effluents being discharged into the water body, or water that was used as a coolant in nuclear power plants. This water is significantly hotter than the water into which it is discharged, which can raise the overall temperature of the water body. Temperature can also be linked to turbidity. Since the amount of light absorbed increases as the water darkens, the temperature will increase.

## **How do we measure temperature?**

Many simple thermometers use thermistor technology. The thermistor is a semiconductor device whose resistance varies as a function of temperature. As the temperature rises, the resistance decreases. This resistance measured by the thermistor is then converted to a value

displayed on the Celsius or Fahrenheit scale. Thermistor sensors are suitable for a temperature range of  $-50^{\circ}$  to  $150^{\circ}$  C ( $-58^{\circ}$  to  $302^{\circ}$  F).

## Temperature Calibration

Many meters are factory calibrated for temperature readings. It is good practice to check at least once a year, in a laboratory setting, that your temperature sensor is working properly.

Conductivity (EC)/ Total Dissolved Solids (TDS)

## What is conductivity?

*Electrical conductivity (EC)* measures how well a substance can transmit an electrical current. Small charged particles, called ions, help to carry the electrical charge through a substance. These ions can be positively or negatively charged. The more ions available, the higher the conductivity; fewer ions would result in lower conductivity. EC is typically reported in milliSiemens per centimeter (mS/cm).

Total dissolved solids (TDS) are the number of dissolved substances in solution. This measurement reads all the dissolved inorganic and organic substances in a liquid. Results from this reading are displayed as milligrams per liter (mg/L), parts per million (ppm), grams per liter (g/L), or parts per thousand (ppt).

## Why is measuring conductivity important?

Electrical conductivity (EC) is another way to evaluate water quality, since the increased presence of total dissolved solids (TDS), as expressed by EC, may be an indicator of pollutants. EC can be impacted by carbonates from limestone, man-made point source pollutants such as sewage treatment plants, or man-made non-point source pollutants such as septic systems or agricultural runoff.

High concentrations of TDS can lower water quality and cause water balance problems for individual organisms. On the other hand, low concentrations may limit the growth of aquatic life. Some of the effects discussed for the acidity and carbon dioxide parameters have relevance for EC, such as its negative impact on photosynthesis. This is because increased solids make water murkier, which slows down the rate of photosynthesis. EC provides an indication of total dissolved solids, of which total dissolved salts are a component. If the level of salts in TDS is high, this could also contribute to the acidity of the water. However, if the level of carbonates in TDS is high, this could contribute to an increase in alkalinity, which helps protect against acidity changes. This is a good example of the interrelationships between water quality parameters.

Acceptable levels of EC in rivers and streams vary depending on the type of dissolved solids present and this determines the use of the stream, such as for fishing, swimming, or as a source of drinking water.

The combination between TDS and Total Solids is important to understand. Total Solids refers to all solid matter that is either suspended or dissolved in water. Dissolved Solids are not visible in water, since through being dissolved they have become part of the solution.

TDS is the measurement of the water-soluble substances that are in a water sample. In a sample solution collected from a river, these dissolved substances are called solutes, and water is called the solvent.

### **How do we measure conductivity?**

The best way to measure conductivity is with the use of an EC meter. Two electrodes with an applied AC voltage are placed in the solution. This creates a current dependent upon the conductive nature of the solution. The meter reads this current and displays in either conductivity (EC) or ppm (TDS).

### **Conductivity Calibration in the Field**

It is important to calibrate for conductivity before sample. This is because oily coatings and biological contaminants can actually change the apparent cell geometry, resulting in a shift in cell constant. Before performing a conductivity calibration, always inspect the EC sensor for debris or blockages.

Most meters are calibrated against a single standard which is near the specific conductance of the environmental sample. A second standard can be used to check the linearity of the instrument in the range of measurements.

#### **Calibration Procedure**

1. Fill a beaker with enough standard to cover the electrode junction (about 75 mL in a 100 mL beaker). Pour additional standard into a second beaker to be used to rinse the sensor.
2. Place the electrode into the rinse beaker and ensure that the EC sensor channels are filled with fresh standard by raising and lowering the beaker a few times.
3. Place the probe into the calibration beaker and tap to dislodge any trapped bubbles.
4. Confirm the calibration point when the reading is stable, or when the digits do not change for at least 5 seconds. (Some meters require you to enter the conductivity standard value).
5. The calibration is complete. Rinse the probe with deionized water and store the probe according to manufacturer's instructions.

#### **Dissolved Oxygen (DO)**

### **What is dissolved oxygen?**

The concentration of dissolved oxygen (DO) in water is extremely important in nature as well as in man's environment. In the oceans, lakes, rivers, and other surface water bodies, dissolved oxygen is essential to the growth and development of aquatic life. Without oxygen, the water can become toxic due to the anaerobic decaying of organic matter. In an industrial environment, water must contain at least 2 mg/L of oxygen to protect water pipes from corrosion. However, boiler system water, in many cases, cannot contain greater than 10 mg/L of oxygen.

### **Why is dissolved oxygen important?**

DO levels can help indicate the relative health of a water body. If DO levels are normal or high, the water is a good environment for a variety of aquatic life to flourish. If DO levels are low, it may indicate the presence of pollutants in the water. Some aquatic life can exist in water with a wide range of DO, but others cannot survive in a low DO environment.

DO measurements are expected to have large fluctuations if the water has significant plant life. This is due to the photosynthesis process. Since there is less photosynthetic activity at night, when light is not present, plants and animals in the water consume oxygen through respiration, but not as much oxygen is produced at the same time. As a result, DO levels in early morning are lower compared to other times of day. Once photosynthesis begins, DO levels will rise. This is a good example of the benefits of measuring parameters at various times throughout the day. If only a pre-dawn DO measurement is taken, an inaccurate conclusion may be drawn regarding the healthiness of the water.

While DO levels are partially influenced by photosynthetic activity, a large source of DO is from atmospheric oxygen mixing with water. This happens in larger amounts if the water is turbulent. The turbulence increases the surface area of the water, so atmospheric oxygen can mix with it more easily. Air has an oxygen concentration that is over 20 times higher than oxygen concentration in water. This concentration difference results in atmospheric oxygen dissolving in water when the two meet. If there is more water surface at this interface, then more oxygen from the air will be absorbed.

Other factors that influence DO levels are temperature and runoff water. Oxygen dissolves more easily in cold water, and cold water has the ability to hold larger levels of gases than warm water, so the DO level decreases as the water gets warmer. Runoff can include natural organic wastes or man-made pollutants; in both cases, organisms in the water must use oxygen in the process of decomposing these pollutants. Also, organic wastes can lead to growth of aquatic vegetation. When plants die at the end of the growing season, heavy oxygen consumption takes place as they are decomposed.

## **How do we measure Dissolved Oxygen?**

Dissolved oxygen concentrations are most often reported in units of milligrams of gas per liter of water, mg/L. (The unit mg/L is equivalent to parts per million = ppm). Measurements are usually taken in water using a DO probe and meter.

It is important to measure DO levels at various times of the day, and at various water depths. These measurements will give an overall picture of the DO levels in the water body being investigated. As with all water quality parameters, these levels must be monitored over time. This will yield a quantity of data points so trends can be identified and evaluated.

## **Dissolved Oxygen Calibration in the Field**

Dissolved oxygen (DO) content in water is measured using an electrode with a membrane. Unfortunately, brushes or other cleaning objects may damage the membrane so replacing the membrane cap and electrolyte is the best way to perform periodic maintenance. Although it may be easier to calibrate the D.O. sensor prior to going out in the field, it is best to calibrate at your sampling site because differences altitude and barometric pressure between the calibration and measurement site can result in errors. Be sure to verify that the barometric pressure, conductivity, and temperature readings are correct.

## Calibration Procedure (100%)

1. Fill a calibration beaker with water (alternatively, a wet sponge or a wet paper towel can be placed on the bottom of the DO calibration container).
2. Loosely fit the probe into the calibration beaker to prevent the escape of moisture.  
*\*Be sure not to get your DO sensor wet as the evaporation of moisture on the temperature sensor or DO probe may influence the readings during calibration.*
3. Allow the container to become saturated with water vapor (approximately 10 to 15 minutes). *\*During this time, turn on the instrument to allow the DO probe to warm-up.*
4. Confirm the calibration point when the reading is stable, or when the digits do not change for at least 5 seconds.
5. The calibration is complete. Rinse the probe with deionized water and store the probe according to manufacturer's instructions.

## References.

Aas, W. and A. Semb. 2001. Standardisation of Methods for Long-term monitoring. *Water, Air, and Soil Pollution* 130:1595-1600.

American Water Works Association (AWWA). 1990. *Water Quality and Treatment: A Handbook of Community Water Supplies*. 4th ed. McGraw-Hill.

APHA (American Public Health Association, American Water Works Association, Water for the Environment Foundation). 2000. *Standard Methods for the Examination of Water and Wastewater*. 20th edition. Editors: L. Clesceri, A. Greenberg, and A. Eaton. , Washington, DC.

ASTM. 2000. *Water: Inorganic Analytes*. Volume 11.01. American Society for Testing and Materials, Conshohocken, PA. Bartholomay, R.C., Knobel, L.L., and Rousseau, J.P., 2003, *Field methods and Quality-Assurance Plan for quality-of-water activities*, U.S. Geological Survey, Idaho National Engineering and Environmental Laboratory, Idaho: U.S. Geological Survey Open-File Report 03-42 (DOE/ID-22182), 45 p.

Battaglin, W.A., Furlong, E.T., Burkhardt, M.R., and Peter, C.J., 2000, Occurrence of sulfonyleurea, sulfonamide, imidazolinone, and other herbicides in rivers, reservoirs and ground water in the Midwestern United States, 1998: *The Science of the Total Environment*, v. 248, no. 2-3, p. 123-133. Brass, H.J., Ardourel, H., Diamond, J., Eaton, A., Keith, L., Peters, C. 2000. Activities of the Interagency Methods and Data Comparability Board. *Proceedings of the American Water Works Association, Water Quality Technology Conference*, Salt Lake City, Utah.

Bushon, Rebecca, 2003, Fecal Indicator Viruses, in> *National field manual for the collection of water-quality data: U.S. Geological Survey Techniques of Water-Resources Investigations*, book 9, chap. A7.

Cantillo, A. Lauenstein, G. 1998. Performance-Based Quality Assurance: The NOAA National Status and Trends Program Experience. In: *Proceedings of the NWQMC National Monitoring Conference, Monitoring: Critical Foundations to Protect Our Waters*. U.S. Environmental Protection Agency, Washington, DC. pp. 63-74.

Crumbling, D. M. 2001. Current Perspectives in Site Remediation Monitoring, EPA 542-R-01-014, October 2001.

D. Yaron. Salinity in Irrigation and Water Resources. Marcel Dekker, Inc., NY., 1981.

Dobson, J., M. Garner, B. Miller, N. Jessup, and R. Toft. 1999. An approach to the assessment of the quality of environmental data. *J. Environ. Monitoring* 1:91-95.

Eaton, A. and Diamond, J. 1999. Reservoir dogs and performance-based systems. *Environ. Testing and Analysis*, 8: 18-19.

GAO. 2001. Environmental Protection: Wider Use of Advanced Technologies can Improve Emissions Monitoring. U.S. General Accounting Office, Report GAO-01-313, Washington, DC.

G.M. Friedman, J.E. Sanders, and D.C. Kopaska-Merkel. Principle of Sedimentary Deposits. Macmillan Publishing Company, NY., 1992.

Grumbly, T. 1994. Institutionalizing the Data Quality Objectives Process for EM's Environmental Data Collection Activities. Memorandum September 7, 1994, United States Department of Energy, Washington, D. C.

G.W. Harvey. Technical Review of Sediment Criteria, for Consideration for Inclusion in Idaho Water Quality Standards. Idaho Dept. of Health and Welfare, Water Quality Bureau, Boise, ID., 1989.

Heinz CSEE. 2002. The state of the nation's ecosystems: measuring the lands, waters, and living resources for the United States. H. J. Heinz III Center for Science, Economics, and the Environment. Cambridge University Press, Cambridge, UK.

ITFM. 1995a. The Strategy for Improving Water Quality Monitoring in the U.S. Report #OFR95-742, U.S. Geological Survey, Reston, VA. ITFM. 1995b. Performance-based

approach to water quality monitoring. In: Strategy for Improving Water Quality Monitoring in the U.S., Appendix M, Report #OFR95-742, Interagency Task Force on Monitoring Water Quality, U.S. Geological Survey, Reston, VA.

J.H. Sherrard, D.R. Moore, and T.A. Dillaha. Total dissolved solids: determinations, sources, effects, and removal. *Journal of Environmental Education* 18(2):19-24, 1987.

Keith, L. 1993. Principles of environmental sampling. ACS Professional Reference Book, American Chemical Society, 458pp.

Koterba, M.T., Wilde, F.D., and Lapham, W.W. 1995. Ground-Water Data-Collection Protocols and Procedures for the National Water-Quality Assessment Program: Collection and Documentation of Water-Quality Samples and Related Data. OFR 95-399, U.S. Geological Survey, Reston, VA.

Lenz, B.N. and M. Miller. 1996. Comparison of Aquatic Macroinvertebrate Samples Collected Using Different Field Methods. USGS Fact Sheet FS-216-96, <http://wi.water.usgs.gov/pubs/FS-216-96/>, U.S. Geological Survey, Reston, VA.

MDCB (Methods and Data Comparability Board). 2002. Method Verification Within a Performance-Based System Framework: Pilot Study Using Chemical Oxygen Demand (COD) Methods. <http://wi.water.usgs.gov/methodsboard/reston/codpilot.htm>

M. Sittig, Handbook of Toxic and Hazardous Chemicals. Noyes Publications. Park Ridge, N.J., 1981.

NELAC. 2000. Chapter 5: Quality Systems Standard. National Environmental Laboratory Conference, [http://www.epa.gov/ttn/nelac/standard/5qs\\_14-0.pdf](http://www.epa.gov/ttn/nelac/standard/5qs_14-0.pdf). June 28, 2000.

North Carolina Administrative Code Section T15A NCAC 02B .0100 & .0200, amended 2002. "RedBook" Surface Water and Wetlands Standards. North Carolina Department of Environment and Natural Resources, Division of Water Quality. Accessed: <http://h2o.enr.state.nc.us/admin/rules/rb010102.pdf>.

NRC (National Research Council). 1995. Review of EPA's environmental monitoring and assessment program: overall evaluation. National Academy Press, Washington, D.C.

NWQMC-Methods Board. 2001. Towards a definition of a performance-based approach to laboratory methods. Methods and Data Comparability Board, National Water Quality Monitoring Council, Technical Report 01-02, US Geological Survey, Water Information Office, Reston, VA.

NWQMC-Methods Board. 2002a. Data Elements for Reporting Water Quality Results of Chemical and Microbiological Analytes. National Water quality Monitoring Council.

NWQMC-Methods Board. 2002b. Accreditation of Federal Laboratories for Water Quality Monitoring. National Water Quality Monitoring Council, February 14, 2002.

O. Postolache, P.M. Girão, J.M. Dias Pereira, H.G. Ramos. "Multibeam Optical System and Neural Processing for Turbidity Measurement", IEEE Sensors Journal, Vol. 7, No. 5, pp. 677 - 684, May, 2007.

Pirkey, K.D. and Glodt, S.R. 2003. Quality Control at the U.S. Geological Survey National Water Quality Laboratory: USGS Fact Sheet 026-98, [http://nwql.usgs.gov/Public/pubs/QC\\_Fact/tet.html](http://nwql.usgs.gov/Public/pubs/QC_Fact/tet.html)

Pritt, J.W. and Raese, J.W. 1995. Quality Assurance/Quality Control Manual—National Water Quality Laboratory: U.S. Geological Survey Open-File Report 95-443, 35 p.

USDOE. 2000. Guide for developing data quality objective for ecological risk assessment at DOE Oak Ridge Operations Facilities. Report ES/ER/TM-185/R1. US Department of Energy, Oak Ridge National Laboratory, Oak Ridge, TN.

USEPA. 1994. Guidelines establishing test procedures for the analysis of pollutants under the Clean Water Act. Final Rule. U.S. Environmental Protection Agency, 40 CFR Part 136, Federal Register. 59:20 4504.

USEPA. 1996. National Primary Drinking Water Regulations: Monitoring Requirements for Public Drinking Water Supplies: Final Rule. U.S. Environmental Protection Agency, 40 CFR Part 141, Federal Register 61(94): 24353-24388.

USEPA. 1997a. Ecological Risk Assessment Guidance for Superfund: Process for Designing and Conducting Ecological Risk Assessments. EPA 540-R-97-006. U.S. Environmental Protection Agency Environmental Response Team, Edison, NJ.

USEPA. 1997 b. Streamlining EPA's Test Methods Approval Program. EPA-821-F-97- 001, U.S. Environmental Protection Agency, Office of Water, Washington, DC.

USEPA. 1998. Synopsis of clarifications to certain Update III SW-846 methods. July, 1998. Office of Solid Waste, Washington, DC.

USEPA. 1999. Protocol for EPA approval of Alternate Test Procedures for Organic and Inorganic Analytes in Wastewater and Drinking water. EPA-821-B-98-002, Office of Water, U.S. Environmental Protection Agency, Washington, DC.

USEPA. 2000. Guidance for the Data Quality Objectives Process (EPA QA/G-4), EPA/600/R-96/055. Quality Assurance Management Staff, Office of Environmental Information. U.S. Environmental Protection Agency, Washington DC.  
<http://www.epa.gov/quality/qs-docs/g4-final.pdf>

USEPA. 2002. Unregulated Contaminant Monitoring Regulation: Approval of Analytical Method for Aeromonas; National Primary and Secondary Drinking Water Regulations: Approval of Analytical Methods for Chemical and Microbiological Contaminants: Final

R.E. Carlson, "The Secchi disk and the volunteer monitor". LakeLine. 15(1): 28-29, 35-37, 1995.

R.E. Carlson and J. Simpson, North American Lake Management Society, "A Coordinator's Guide to Volunteer Lake Monitoring Methods", 96 pp, 1996.

R.L. Smith. Ecology and Field Biology. 4th ed. Harper Collins Publishers, NY., 1990.

Rule.40 CFR Part 141,U.S. Environmental Protection Agency, Federal Register67: (209): 65888-65902.

T. Dunne, and L.B. Leopold. Water in Environmental Planning. W.H. Freeman and Company, NY., 1978.

Ward, R.C. 1996. Water quality monitoring: where's the beef? American Water Resources Association. 32:673-680.

Wilde, F.D., Radtke, D.B., Gibs, Jacob, and R.T. Iwatsubo. 1998. Preparations for Water Sampling. In National Field Manual for the Collection of Water-Quality Data: U.S. Geological Survey Techniques of Water-Resources Investigations, book 9, chap. A1, 42 p. ([http://water.usgs.gov/owq/FieldManual/chapter1/html/Ch1\\_contents.html](http://water.usgs.gov/owq/FieldManual/chapter1/html/Ch1_contents.html))

Wilde, F.D., Radtke, D.B., Gibs, Jacob, and R.T. Iwatsubo. 1999. Collection of Water Samples. In National Field Manual for the Collection of Water-Quality Data: U.S. Geological Survey Techniques of Water-Resources Investigations, book 9, chap. A4, 151 p. ([http://water.usgs.gov/owq/FieldManual/chapter4/html/Ch4\\_contents.html](http://water.usgs.gov/owq/FieldManual/chapter4/html/Ch4_contents.html))

Wiley, M.J., P.W. Seelbach, K. Wehrly, and J.S. Martin. 2003. Regional ecological normalization using linear models: a meta- method for scaling stream assessment indicators. In: T. P. Simon (ed) Biological Response signatures, pp.201-224, CRC Press, Boca Raton, FL.

World Bank Group. 2007. Access to Safe Water Map. Available at: <http://www.worldbank.org/depweb/english/modules/environm/water/map1.html>